

## Supporting Information

For

**Guest and Subunit Exchange in Self-Assembled Ionophores**

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Figure 1. The <sup>1</sup>H NMR spectrum of cation-free isoG **1** is concentration dependent.

Figure 2. Representative <sup>1</sup>H NMR spectrum of an (isoG **1**)<sub>10</sub>-M<sup>+</sup>BPh<sub>4</sub><sup>-</sup> decamer.

Figure 3. Molar ratio <sup>1</sup>H NMR experiments for alkali cation complexation by isoG **1**.

Figure 4. Stack plot of <sup>133</sup>Cs NMR spectra.

Figure 5. Determination of the Cs<sup>+</sup> exchange rate.

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**Cesium-133 NMR Spectroscopy.** Cesium-133 NMR spectra were obtained on a Bruker DRX-500 MHz spectrometer (65.6 MHz for  $^{133}\text{Cs}$ ). The  $^{133}\text{Cs}$  chemical shifts were referenced to an external solution of 0.50 M CsI in  $\text{D}_2\text{O}$  at 298 K. Samples for  $^{133}\text{Cs}$  NMR were prepared by addition of isoG **1** into solutions of  $\text{Cs}^+\text{BPh}_4^-$  (of known concentration in either  $\text{CD}_3\text{CN}$  or 50%-50%  $\text{CD}_3\text{CN-CDCl}_3$ ).  $\text{H-1}$  NMR determined the stoichiometry, as illustrated in Figure 2 of the Supporting Information.

**2D  $^1\text{H-}^1\text{H}$  EXSY Measurements.** Samples for the 2D EXSY NMR experiments were prepared by mixing the metal-free ligand isoG **1** with  $(\text{isoG } \mathbf{1})_{10}\text{-M}^+\text{BPh}_4^-$  decamer complexes. The  $(\text{isoG } \mathbf{1})_{10}\text{-M}^+\text{BPh}_4^-$  decamer complexes were prepared in  $\text{CDCl}_3$  and characterized by  $^1\text{H}$  NMR, then metal-free ligand isoG **1** was added to give a ratio of isoG **1**:  $(\text{isoG } \mathbf{1})_{10}\text{-M}^+ = 10:1$ . Similarly, samples were also prepared in the mixed solvent of 50%-50%  $\text{CD}_3\text{CN-CDCl}_3$ . These samples contained 40 mM of isoG **1** and 2.0 mM of  $\text{Cs}^+$ , giving a ratio of isoG **1**:  $(\text{isoG } \mathbf{1})_{10}\text{-M}^+ = 10:1$ .

Two-dimensional EXSY NMR spectra were recorded on a Bruker DRX-500 MHz spectrometer at various temperatures. The typical NOESY pulse sequence was used, with mixing times varying from 0.1 s to 1.3 s, and with relaxation time of 4.0. The sweep width in both dimensions was 7500 Hz (at 500 MHz). A total of 512  $t_1$  values were used, and 2048-point FIDs were acquired in the  $t_2$  dimension. A line broadening of 2 Hz in  $F_2$  dimension and 7 Hz in  $F_1$  dimension was imposed prior to Fourier transformation. The total time that was required for each experiment was 18 h.

Sizable off-diagonal peaks were observed, indicating exchange between decamer and uncomplexed isoG **1**. The peak intensity,  $I_{ij}$  ( $i, j = \text{monomer, complex}$ ) was measured using the WinNMR program (Bruker). The relationship between the 2D peak intensities ( $I_{ij}$ ) at some specified mixing time ( $t_m$ ) and the exchange rate constants is given by following equation:

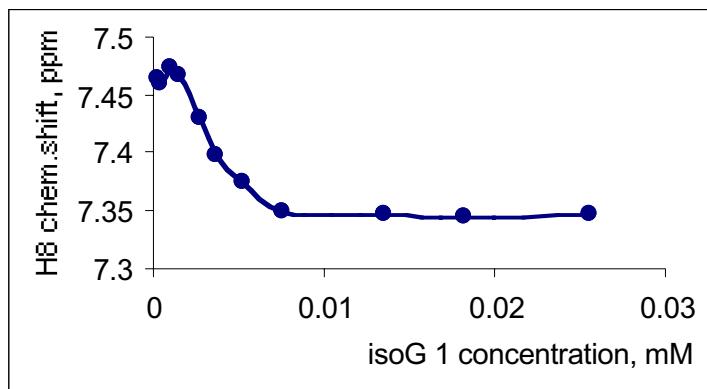
$$I_{ij}(t_m) = (e^{-\mathbf{R} t_m})_{ij} M_j^0$$

$M_j^0$  is the equilibrium magnetization of the nuclei in site  $j$  and  $\mathbf{R}$  has off-diagonal elements  $\mathbf{R}_{ij} = -k_{ji}$ , where  $k_{ji}$  is the first order rate constant for chemical exchange from site  $j$  to site  $i$ . The rate constants  $k_{ij}$  and  $k_{ji}$  were calculated using the matrix method (see Perrin, C. L.; Gipe, R. K. *J. Am. Chem. Soc.* **1984**, *106*, 4036-4038; and Perrin, C. L.; Dwyer, T. J. *Chem. Rev.* **1990**, *90*, 935-967):

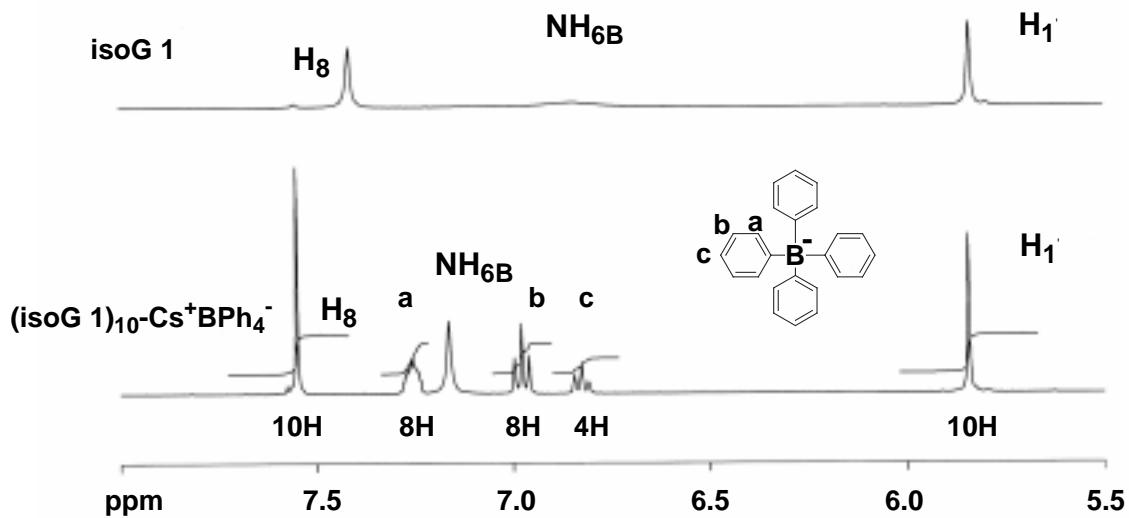
$$\mathbf{R} = -\tau_m^{-1} \ln A = -\tau_m^{-1} X(\ln \Lambda)X^{-1}$$

Where  $\mathbf{R}_{ij} = -k_{ji}$ ,  $A_{ij} = I_{ij}(\tau_m / M_j^0)$  and  $X$  is the square matrix of eigenvectors of  $A$ , such that  $X^{-1}AX = \Lambda = \text{diag}(\lambda_i)$  and  $\ln \Lambda = \text{diag}(\ln \lambda_i)$ , with  $\lambda_i$  the  $i$ th eigenvalue of  $A$ .

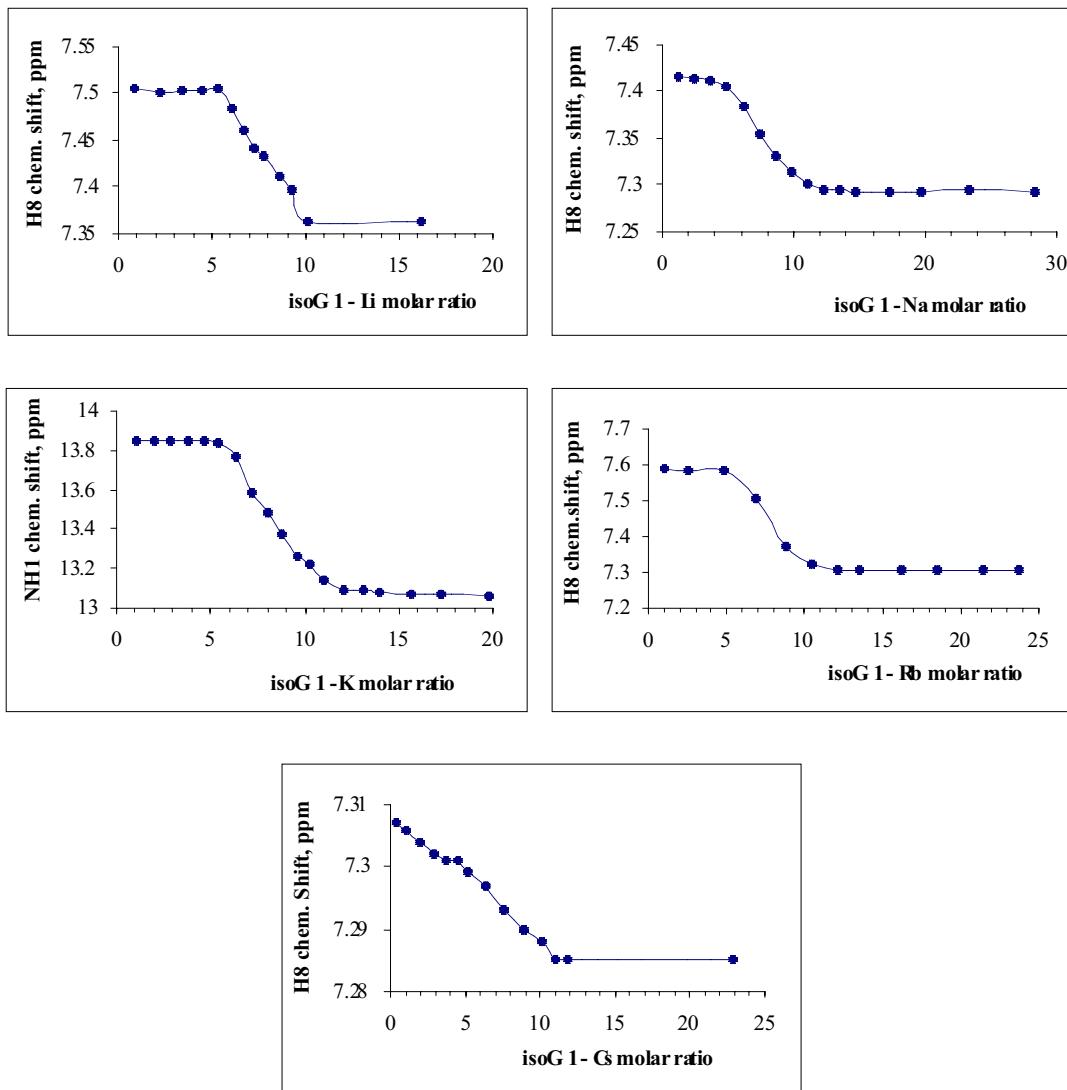
Calculations were done using MathCAD 7 software (MathSoft Inc.). The mean lifetime of exchange  $\tau$  was calculated from the relationship to the measured first-order rate constants  $k_{ij}$  and  $k_{ji}$ ,  $\tau = 1/k$  ( $k$  is the average exchange rate constant,  $k = (k_{ij} + k_{ji})/2$ ).



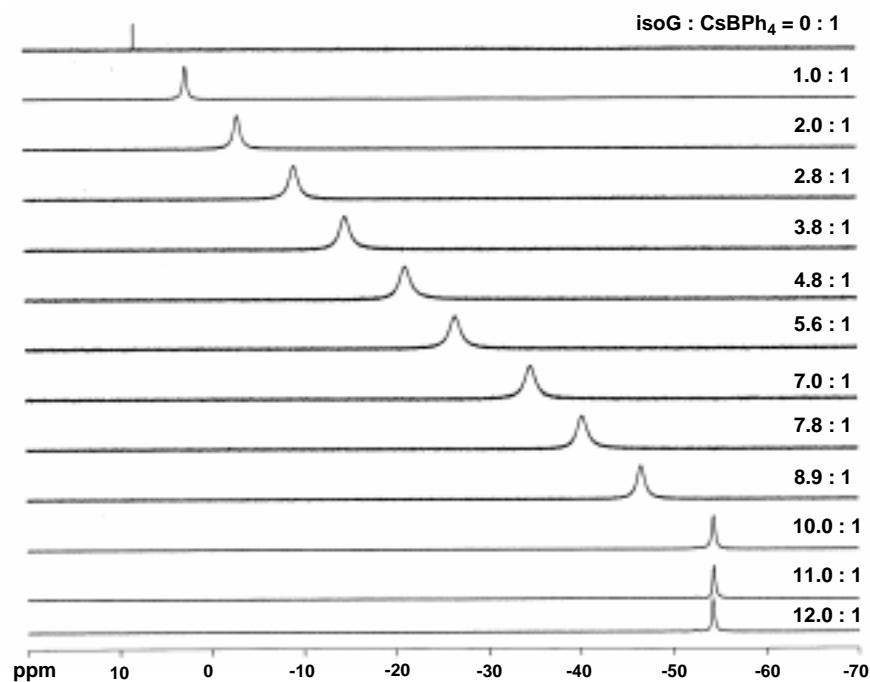
**Fig. 1.** The  $^1\text{H}$  NMR spectrum of cation-free isoG **1** is concentration dependent. This plot illustrates the chemical shift of the aromatic H8 proton in a 1:1  $\text{CDCl}_3\text{-CD}_3\text{CN}$  solution as a function of the concentration of isoG **1**. We conclude that the observed changes in the  $^1\text{H}$  NMR spectrum are due to non-specific self-association of cation-free isoG **1**. These NMR results are consistent with isothermal titration calorimetry and VPO data described in the text of the paper



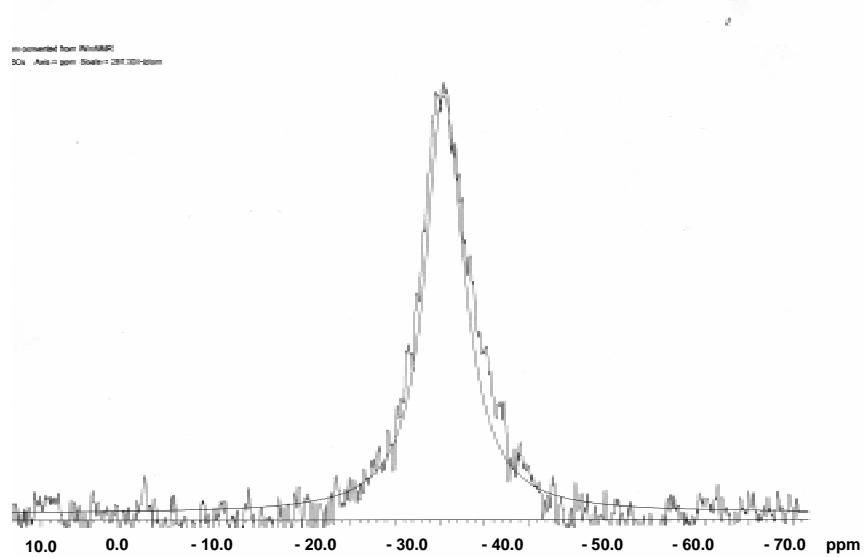
**Fig. 2.** Representative  $^1\text{H}$  NMR spectra of metal-free isoG **1** (top) and  $(\text{isoG 1})_{10}\text{-Cs}^+\text{BPh}_4^-$  decamer. The decamer sample was generated by titration of isoG **1** into a solution of  $\text{Cs}^+\text{Ph}_4\text{B}^-$  in  $\text{CD}_3\text{CN}$ . The isoG **1** to  $\text{Cs}^+\text{Ph}_4\text{B}^-$  stoichiometry in complex was determined by comparing the integration of the  $\text{BPh}_4^-$  signals (the ortho-, meta-, and para-protons) with the integration of signals corresponding to the complexed isoG **1**.



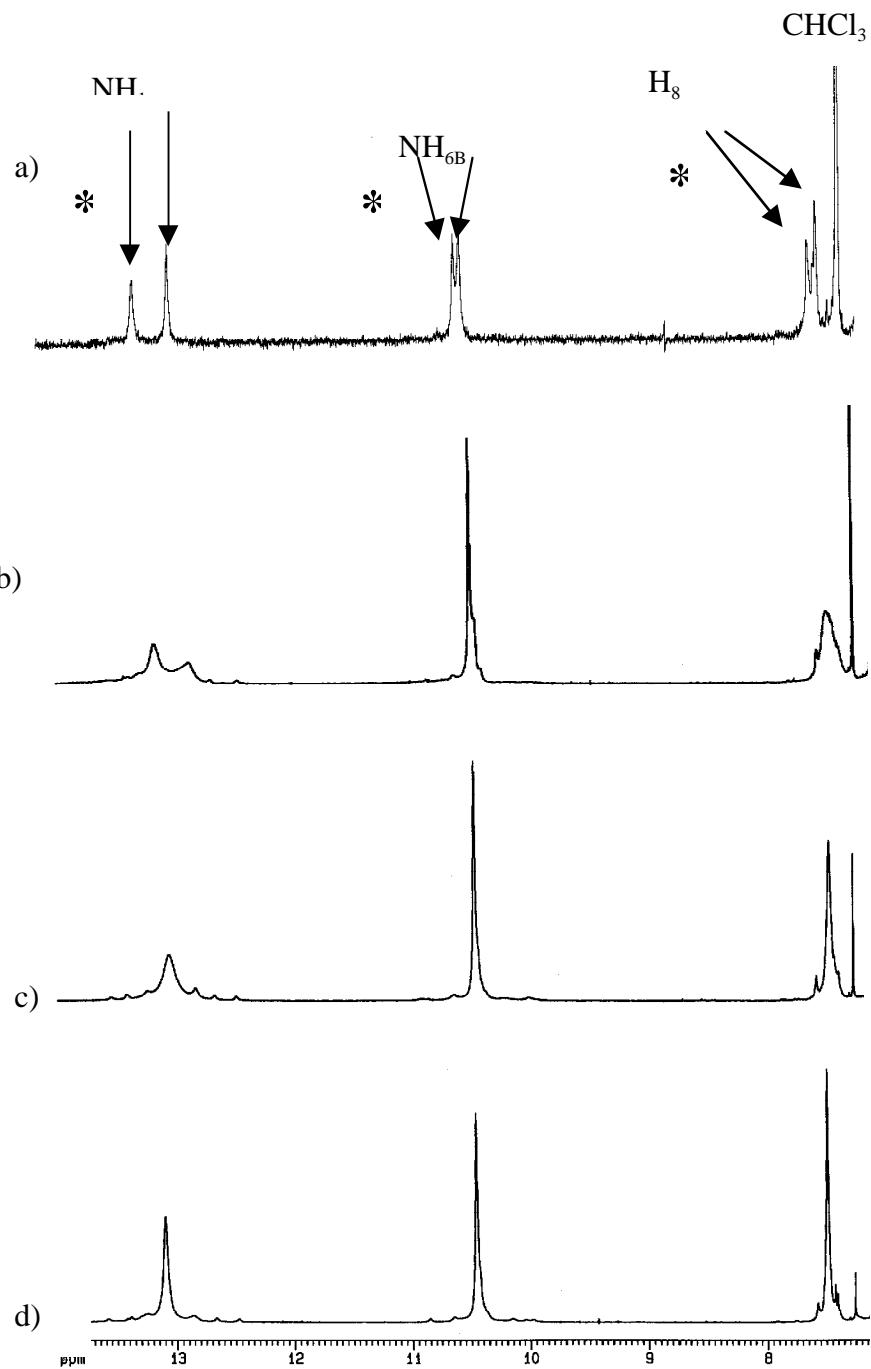
**Fig. 3.** Molar ratio  $^1\text{H}$  NMR experiments for alkali cation complexation by isoG **1**. Experiments were done in a mixed 1:1  $\text{CDCl}_3/\text{CD}_3\text{CN}$  solvent using alkali tetraphenylborate salts. Relative stoichiometries were determined by integration of the tetraphenylborate and isoG NMR peaks, as described in Supplemental Figure #2. The salt concentrations ranged between 2.5 mM and 11.3 mM. All alkali metal cations, except  $\text{Cs}^+$ , show two inflection points corresponding to a 5:1 pentamer and 10:1 decamer complex. We observed only one inflection point in the isoG- $\text{Cs}^+$  experiment, corresponding to 10:1 decamer stoichiometry



**Fig. 4.** Stack plot of <sup>133</sup>Cs NMR spectra. Titration of isoG **1** into a CsBPh<sub>4</sub> solution (10 mM) in CD<sub>3</sub>CN demonstrated: (i) fast exchange of Cs<sup>+</sup> between the solvated and bound states; and (ii) the complex formed is a decamer, (isoG **1**)<sub>10</sub>-Cs<sup>+</sup>BPh<sub>4</sub><sup>-</sup>.



**Fig. 5.** Determination of the  $\text{Cs}^+$  exchange rate. Experimental  $^{133}\text{Cs}$  NMR spectra were imported into the g-NMR 4 program (Cherwell Scientific, Palo Alto, CA, 94303) for computer simulation of  $\text{Cs}^+$  exchange and for calculation of the mean lifetime ( $\tau$ ) of  $\text{Cs}^+$  in the decamer complex  $(\text{isoG } \mathbf{1})_{10}\text{-Cs}^+\text{BPh}_4^-$ . Simulated data were iteratively fit to the experimental data. The experimental and simulated data for a sample containing a solution of  $(\text{isoG } \mathbf{1})_{10}\text{-Cs}^+\text{Ph}_4\text{B}^-$  (1 mM) and  $\text{Cs}^+\text{Ph}_4\text{B}^-$  (1 mM) in 50%  $\text{CDCl}_3$ -50%  $\text{CD}_3\text{CN}$  at 25 °C is shown. Line-shape analysis provided a mean lifetime of  $\tau=0.061$  ms for bound  $\text{Cs}^+$ .



**Fig. 6.** NMR dilution experiments show exchange between the pentamer and decamer. In addition to being cation dependent, (see reference 12 in the paper), pentamer-decamer exchange was also concentration-dependant, indicating that exchange proceeds via a bimolecular mechanism. This Figure shows a region of the  $^1\text{H}$  NMR spectra for a 1:2 mixture of  $(\text{isoG } \mathbf{1})_{10}\text{-Li}^+$  and  $(\text{isoG } \mathbf{1})_5\text{-Li}^+$  in a 1:1  $\text{CDCl}_3\text{:CD}_3\text{CN}$  solvent mix. Arrows marked with asterisk indicate the pentamer resonances. The total concentration of isoG **1** is a) 1.3 mM; b) 9.4 mM; c) 21.6 mM; d) 54.9 mM.